Biocontrol of Anopheles gambiae larvae using fresh ripe and unripe fruit extracts of Capsicum frutescens var. baccatum

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Abstract

Purpose: This study investigated the biocontrol of *Anopheles gambiae* larvae using fresh ripe and unripe fruit extracts of *Capsicum frutescens* var. *baccatum*. **Materials and Methods:** Ripe and unripe fruit extracts of *C. frutescens* var. *baccatum* were tested against *A. gambiae* for 24 h. The *A. gambiae* used for the study was obtained from the wild and identified following standard procedure. **Results and Discussion:** Results of the bioassay showed that 0, 50, 100, 150, 200, and 250 ppm concentration, the percentage mortality was 0.00%, 16.67%, 38.33%, 46.67%, 70.00%, and 86.67%, respectively, for the unripe extracts, and 0%, 10.00%, 26.67%, 40.00%, 40.00%, and 66.67%, respectively, for the ripe extract. Analysis of variance showed that there was a significant difference at P < 0.05 across the concentrations. Between unripe and ripe extracts, there were no significant variations (P > 0.05) at 50, 100, and 150 ppm, but significant variations (P < 0.05) exist at 200 and 250 ppm. The LC₅₀value was 121.65 ppm for the unripe extract and 196.61 ppm for the ripe extracts. **Conclusion and Recommendation:** From the result of this study, there is the need for research to focus on the isolation and purification bioactive compounds that make the pepper confer activity against of larvae of *A. gambiae*.

Key words: Capsicum frutescens fruit, extracts, medicinal plants, mosquito, vector

INTRODUCTION

ithin the past decades, the use of pesticides has increased in agricultural activities and household setting. The use of chemical was originally seen as a solution to many pests. Pesticides are classified based on their chemical compositions as well as the target organisms. On targets organisms, the notable classification includes insecticides, rodenticides, fungicides, herbicides, and fumigants. The use of these pesticides can cause pollution depending on the dose/concentration. Typically, these pesticides have the tendency to drift and they could end up in the aquatic ecosystem through spill during spraying, runoff of remain of cans, and/or farmland close to the aquatic ecosystem. On the aquatic ecosystem, studies have reported that many pesticides could affect hematological, enzymes, metabolites, electrolytes, behavioral, and mortality rate of fishes.^[1-4]

Insecticides are substances that are used to eliminate insects specifically. Most commercially available insecticides have the tendency to kill both insects that undergo complete (mosquito) and incomplete (cockroaches) metamorphosis. As such, insecticides most have the tendency to act on the eggs, larvae, pupa, and adult of insects such as mosquito as well as egg, nymph, and adult of cockroaches. In agricultural activities, insecticides are very useful for increased productivity, especially in areas that insects threaten agricultural produce. Again on human health, insecticides are used to control some vector-borne diseases.

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Received: 26-03-2019 **Revised:** 05-07-2019 **Accepted:** 26-07-2019 Different brands of insecticides are available in the market today. Like most pesticides, many of the insecticides are harmful to humans, and some have the tendency to concentrate along the food chain. Many insecticides can be grouped based on their activity through systemic (insecticides with long-term activity) and contact (insecticides with short-term effect or no residual activity). Before now, many of the insecticides available in the market are produced with inorganic compounds and due to technology organic-based insecticides were developed and it works mainly by contact.

Due to the toxicity of inorganic-based insecticides, research into the production of natural products for the control of insects has been carried out and a major breakthrough had been achieved. To these effects, many plants have been found to have insecticidal potential. Authors have reported that the pharmacological characteristics of the plant are associated with their bioactive compounds.^[5-12] Some of the bioinsecticides that have been well established are pyrethrum and neem.^[13] These natural insecticides are less toxic and could biodegrade compare the inorganic-based insecticides.

In Sub-Saharan Africa, malaria is among the major vectorborne diseases. In 2010, about 81% of global malaria infection occurred in Africa.^[14] The reported also indicated that about 50% of world malaria cases occurred in Nigeria, Democratic Republic of Congo, Ethiopia, and Uganda causing millions of dead per annum. Besides malaria, mosquito also transmits filariasis, yellow fever, dengue fever, encephalitis, etc.^[15-17] Among the many genera of mosquito, *Aedes, Culex, Anopheles,* and *Mansonia* are known to transmit important diseases in human and animals.^[15] Specifically, the *Anopheles* mosquito transmits malaria. Of all the species of *Anopheles, Anopheles gambiae* and *Anopheles arabiensis* transmit most malaria cases in Africa.^[15,18,19]

In many regions that malaria is endemic, the genus *Anopheles* is the most important iniquitous dipterans fly that transmits the disease.^[11,12] In addition, *Aedes aegypti* transmits chikungunya, yellow and dengue fevers, while *Culex quinquefasciatus transmits lymphatic filariasis*.^[15]

Based on the safe use of natural insecticides when compared with inorganic-based insecticides, research has well carried out to determine the activities of different plant extract against the mosquito. Several plants have been used to control the larvae of *A. gambiae* using different solvents.^[15] Izah^[11] reported the activities of ethanolic, acetone, and crude extracts of *Capsicum frutescens* var. *minima* fruit against *A. gambiae* larvae. However, information about the biocontrol of *A. gambiae* larvae using fresh ripe and unripe fruit extracts of *C. frutescens* var. *baccatum* is scanty in literature, hence, the need for this study.

MATERIALS AND METHODS

Plant Collection and Extraction

The fruit of fresh ripe and unripe *C. frutescens* var. *baccatum* was obtained from a smallholder farmer in Ndemili, Delta state, Nigeria. About 20 g of the pepper were macerated using pestle and mortar and 100 ml of water was added. The medium was allowed to extract for 48 h. Then, after the pepper was filter pressed using a double muslin cloth. The filtrate was reconstituted with distilled water to different concentration.

Culture of A. gambiae

A. gambiae larvae used for the study was obtained from the wild with the assistant of baits in abandoned plastic containers half-filled with water, cotton wool, and debris. The water was carefully removed from the container containing the bait and syringe without the needle section was used to transfer the larvae. Some of *A. gambiae* larvae obtained were allowed to develop into adults and identified based on the morphology previously presented by Ahmed and Ahmed (2011), and Gimba and Idris (2014).^[20,21] The *A. gambiae* larva was fed with biscuit and yeast at a ratio of 3:1 at room temperature ($27 \pm 3^{\circ}$ C).

Larvicidal Bioassay

The activities of the fruit of fresh ripe and unripe *C. frutescens* var. *baccatum* against larvae of *A. gambiae* were carried out based on the scheme of the WHO^[22] cited by Rathy *et al.*^[23] A concentration of 0, 50, 100, 125, 150, 200, and 250 ppm of the extracts was made with dechlorinated. Then, 20 larvae of *A. gambiae* were introduced into each of the concentration. At 24 h, after the commencement of the experiment, the mortality rate was determined. The larvae that remain motionless and did not respond to prodding with a soft brush were considered dead.

The percentage of mortality was calculated as

Number of dead larva Total number of larva in the experimental group

Statistical Analysis

The percentage mortality between the various concentrations was subjected to one-way analysis of variance at P = 0.05, and the mean value was converted to charts using Microsoft Excel. Where significant variations occurred, Duncan multiple range test statistics was used to determine the source of the variation. Furthermore, *t*-test was used to show variations between ripe and unripe extract for each concentration. The LC₅₀ value was calculated based on probit analysis using Finney's table.^[24] Microsoft Excel Window 2010 was used to

carry out the regression analysis, the probit value against log concentration from the equation of the study was determined and the LC_{50} was calculated by substitution of the probit value of 50 and then finding the antilogarithm of the value.

RESULTS AND DISCUSSION

Figure 1 presents the activities of fresh ripe and unripe fruit extracts of C. frutescens var. baccatum against larvae of A. gambiae at 24 h. At 0, 50, 100, 150, 200, and 250 ppm, the percentage mortality was 0%, 16.67%, 38.33%, 46.67%, 70.00%, and 86.67%, respectively, for the unripe extracts. Statistically, there was a significant difference at P < 0.05across the various concentrations except for 100 and 150 ppm that mean separation showed that there was not significantly different. Percentage mortality was 0%, 10.00%, 26.67%, 40.00%, 40.00%, and 66.67% for 0, 50, 100, 150, 200, and 250 ppm, respectively, for ripe extract. There was a significant difference at P < 0.05 for the various concentrations. Pairwise comparisons between the various comparisons showed that there is no significant difference between 0 and 50 ppm, and between 150 and 200 ppm. Between unripe and ripe extracts, there were no significant variations (P > 0.05) at 50, 100, and 150 ppm, but significant variations (P < 0.05) exist at 200 and 250 ppm [Table 1]. The findings of this study revealed that the stage of the development of the pepper (C. frutescens var. baccatum) can induce varying toxicity in the larvae of A. gambiae. In general, the unripe extract had a superior effect which was relatively higher at 50-150 ppm and statistically different at 200-250 ppm [Figure 1 and Table 1]. The variation in the toxicity could be due to differences in the moisture content of the pepper as well as the concentration of the active ingredients of the pepper that makes it conforms the larvicidal activities.

The LC₅₀ value of *A. gambiae* larvae exposed to unripe fresh fruit extracts of *C. frutescens* var. *baccatum* fruit for 24 h is 121.65 ppm [Figure 2], while the LC₅₀ values of *A. gambiae* larvae exposed to ripe fresh fruit extracts of *C. frutescens* var. *baccatum* fruit for 24 h are 196.61 ppm [Figure 3]. Again the LC₅₀ value showed that the unripe extract had superior activity against the larvae of *A. gambiae*. The relative difference may be associated with the concentration of the active ingredients in each of the paper.^[11,12] Some of the active compounds that

have been reported in different varieties of *C. frutescens* include alkaloids, tannins, steroids, glycosides, saponins,

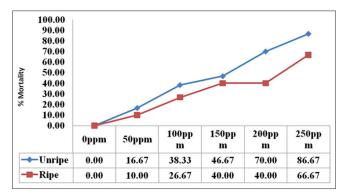


Figure 1: Activities of fresh ripe and unripe fruit extracts of *Capsicum frutescens* var. *baccatum* against larvae of *Anopheles gambiae* at 24 h

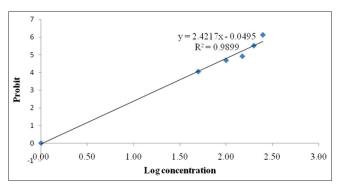


Figure 2: LC₅₀ value of *Anopheles gambiae* larvae exposed to unripe fresh fruit extracts of *Capsicum frutescens* var. *baccatum* fruit for 24 h

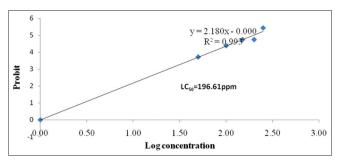


Figure 3: LC₅₀ value of *Anopheles gambiae* larvae exposed to ripe fresh fruit extracts of *Capsicum frutescens* var. *baccatum* fruit for 24 h

Table 1: Comparative activities of fresh ripe and unripe fruit extracts of Capsicum frutescens var. baccatumagainst larvae of Anopheles gambiae at 24 h				
Concentration, ppm	Unripe (<i>n</i> =3)	Ripe (<i>n</i> =3)	<i>t</i> -test	<i>P</i> -value
50	16.67±5.77	10.00±5.00	1.512	0.205
100	38.33±7.64	26.67±7.64	1.871	0.135
150	46.67±2.89	40.00±5.00	2.000	0.116
200	70.00±5.00	40.00±5.00	7.348	0.002
250	86.67±2.89	66.67±7.64	4.243	0.013

Data are expressed as mean±standard deviation

flavonoids, phenol, carbohydrate, protein, reducing sugar, and capsaicin.[25-27] Basically, capsaicin accounts for the pungent smell of the pepper,^[27] while alkaloids may account for its ability to wade of pests.^[28] The LC₅₀ values of this study vary from the LC₅₀ value of 231.59 ppm (A. aegypti) and 300 ppm (Aedes albopictus) for crude extracts and 97.22 ppm (A. aegypti) and 41.74 ppm (A. albopictus) for ethyl acetate fraction of C. frutescens fruit as reported by Alvarez et al.^[29] Youkparigha and Izah^[12] reported LC₅₀ value of 316.96 ppm, 176.20 ppm, and 136.14 ppm for cold water extracts, hot water extracts, and boiled extracts of ginger, Zingiber officinale, respectively, against A. gambiae, a malaria vector. Izah^[11] reported LC₅₀ value of 115.24 ppm, 173.16 ppm, and 265.19 ppm for ethanolic, acetone, and crude extracts of C. frutescens var. minima fruit, respectively, against A. gambiae larvae. The variation may be due to the biochemical/genetic makeup of the different mosquito as well the concentration of active compounds found in the pepper.^[11,12]

CONCLUSION

This study evaluated the activities of fresh ripe and unripe fruit extracts of *C. frutescens* var. *baccatum* against larvae of *A. gambiae* at 24 h. The study found that unripe extract had LC_{50} value of 121.65 ppm being apparently superior compared to the ripe (with LC_{50} value of 196.61 ppm) extracts. The variations may be associated with the moisture content and difference in the phytochemical composition of the pepper that makes it confers larvicidal activity. As such, there is the need for research to focus on isolation and purification of the exact compound that makes the plant to kill the larvae of *A. gambiae*.

REFERENCES

- Inyang IR, Ajimmy R, Izah SC. Organosomatic index and behavioral response of *Heterobranchus bidorsalis* exposed to rhonasate 360sl containing glyphosate (isopropylamine salt glycine). ASIO J Microbiol Food Sci Biotechnol Innov 2017;3:6-14.
- Inyang IR, Izah SC, Johnson DT, Ejomarie OA. Effects of lambda cyhalothrin on some electrolytes and metabolites in organs of *Parpohiocephalus obscurus*. Biotechnol Res 2017;3:6-10.
- Inyang IR, Ollor AO, Izah SC. Effect of diazinon on organosomatic indices and behavioural responses of *Clarias gariepinus* (a Common Niger Delta Wetland Fish). Greener J Biol Sci 2017;7:15-19.
- 4. Inyang IR, Seiyaboh EI, Job UB. Condition factor, organosomatic indices and behavioural abnormalities of *Clarias gariepinus* exposed to lambda cyhalothrin. Greener J Life Sci 2017;4:1-5.
- Izah SC. Some determinant factors of antimicrobial susceptibility pattern of plant extracts. Res Rev Insight 2018;2:1-4.

- Izah SC, Uhunmwangho EJ, Dunga KE. Studies on the synergistic effectiveness of methanolic extract of leaves and roots of *Carica papaya* L. (papaya) against some bacteria pathogens. Int J Complement Altern Med 2018;11:375-8.
- Izah SC, Uhunmwangho EJ, Eledo BO. Medicinal potentials of *Buchholzia coriacea* (wonderful kola). Med Plant Res 2018;8:27-43.
- Izah SC, Uhunmwangho EJ, Etim NG. Antibacterial and synergistic potency of methanolic leaf extracts of *Vernonia amygdalina* L. and *Ocimum gratissimum* L. J Basic Pharmacol Toxicol 2018;2:8-12.
- Izah SC, Uhunmwangho EJ, Dunga KE, Kigigha LT. Synergy of methanolic leave and stem-back extract of *Anacardium occidentale* L. (cashew) against some enteric and superficial bacteria pathogens. MOJ Toxicol 2018;4:209-11.
- Izah SC, Zige DV, Alagoa KJ, Uhunmwangho EJ, Iyamu AO. Antibacterial efficacy of aqueous extract of *Myristica fragrans* (Common Nutmeg). EC Pharmacol Toxicol 2018;6:291-5.
- 11. Izah SC Activities of crude, acetone and ethanolic extracts of *Capsicum frutescens* var. minima fruit against larva of *Anopheles gambiae*. J Environ Treat Tech 2019;7:196-200.
- 12. Youkparigha FO, Izah SC. Larvicidal efficacy of aqueous extracts of *Zingiber officinale* Roscoe (ginger) against malaria vector, *Anopheles gambiae* (*Diptera: Culicidae*). Int J Environ Agric Sci 2019;3.
- 13. Khater HF Prospects of botanical biopesticides in insect pest management. Pharmacologia 2012;3:641-56.
- Nigeria Malaria Fact Sheet; 2011. Available from: http:// www.photos.state.gov/libraries/nigeria/487468/pdfs/ DecemberMalariaFactSheet%202.pdf. [Last accessed on 2017 Jan 12].
- 15. Bassey SE, Izah SC. Nigerian plants with insecticidal potentials against various stages of mosquito development. ASIO J Med Health Sci Res 2017;2:7-18.
- 16. Bassey SE, Izah SC. Some determinant factors of malaria prevalence in Nigeria. J Mosq Res 2017;7:48-58.
- 17. Ndiok EO, Ohimain EI, Izah SC. Incidence of malaria in Type 2 diabetic patients and the effect on the liver: A case study of Bayelsa state. J Mosq Res 2016;6:1-8.
- Hamza AM, Rayah EA, Abukashawa SM. Molecular characterization of mosquitoes of *Anopheles gambiae* species complex (*Diptera: Culicidae*) from Sudan and republic of Southern Sudan. J Mosq Res 2014;4:1-10.
- Owoeye JA, Akawa OB, Akinneye JO, Oladipupo SO, Akomolede OE. Toxicity of three tropical plants to mosquito larvaee, pupae and adults. J Mosq Res 2016;6:1-7.
- Ahmed UA, Ahmed MM. Morphological identification of malaria vectors within *Anopheles* species in parts of Kano state, Nigeria. Bayero J Pure Appl Sci 2011;4:160-3.
- 21. Gimba UN, Idris HS. Morphological identification and distribution of *Anopheles* species in Gwagwalada

town, F.C.T, Nigeria. Int J Environ Sci Toxicol Res 2014;2:210-6.

- 22. World Health Organization. Report of the WHO Information Consultation on the Evaluation on the Testing of Insecticides CTD/WHO PES/IC96. Geneva: World Health Organization; 1996.
- 23. Rathy MC, Sajith U, Harila CC. Larvicidal efficacy of medicinal plant extracts against the vector mosquito *Aedes albopictus*. Int J Mosq Res 2015;2:80-2.
- 24. Finney DJ. Probit Analysis. London, United Kingdom: Cambridge University Press; 1971.
- 25. Bello I, Boboye BE, Akinyosoye FA. Phytochemical screening and antibacterial properties of selected Nigerian long pepper (*Capsicum frutescens*) fruits. Afr J Microbiol Res 2015;9:2067-78.
- 26. Sen N, Paul D, Sinha SN. In vitro antibacterial potential

and phytochemical analysis of three species of chilli plant. J Chem Pharm Res 2016;8:443-7.

- Vinayaka KS, Prashith KT, Nandini KC, Rakshitha MN, Ramya M, Shruthi J, *et al.* Potent insecticidal activity of fruits and leaves of *Capsicum frutescens* (L.) var. longa (*Solanaceae*). Der Pharm Lett 2010;2:172-6.
- 28. Agu GC, Thomas BT. Antibacterial activities of ethanol and aqueous extracts of five Nigerian medicinal plants on some wound pathogens. Nat Sci 2012;10:78-84.
- 29. Alvarez MR, Heralde FM, Quiming NS. Potent larvicidal activities of *Capsicum frutescens* (L.) fruit ethanolic and partially purified extracts against *Aedes aegypti* (L.) and *Aedes albopictus* (S.). Der Pharm Lett 2015;7:94-9.

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